

## PATENT SPECIFICATION

NO DRAWINGS

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## COMPLETE SPECIFICATION

Improvements in or relating to Ingestible Dry Micro-Organism  
preparations and method for producing the same

I, JOHANN CARL PFLÜGER, a German Citizen, trading as the Firm MED FABRIK CHEMISCH-PHARMAZEUTISCHER PRÄPARATE J. CARL PFLÜGER, of 19 to 22, Karlsruher Strasse, Berlin-Halensee, Germany, do hereby declare the invention, for which I pray that a patent may be granted to me, and the method by which it is to be performed, to be particularly described in and by the following statement:—

This invention relates to ingestible dry microorganism preparations and more particularly to dry preparations of living microorganisms such as the bacteria of the normal intestinal flora, and to methods of manufacturing such preparations so that the same remain stable over prolonged periods of time.

Scientific investigations of symbiosis have shown that an intestinal flora is demonstrably necessary for survival of lower animals, and that the symbionts are not made superfluous by compensating nutrition. Symbionts and microorganisms are inseparably connected during the entire life-span and the symbionts furnish growth substances which are absolutely necessary for normal development. The significance of symbionts to animals could be proved experimentally.

Whether the vital necessity of the intestinal flora which was established for the animal world is also of significance to man, was not known. It was only with the introduction of antibiotics therapy that it became possible to study experimentally the significance of the physiologically normal intestinal symbionts as the intestinal flora was destroyed by the antibiotics. Detrimental side effects after use of antibiotics have been increasingly reported in recent years and have furnished adequate evidence that the destruction of a physiologically normal intestinal flora can lead to abnormal conditions of great severity. When it became known that the antibiotics not only kill pathogenic bacteria but also microorganisms which

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are part of the normal human physiology, scientists started investigating the question of the necessity of the intestinal flora in humans.

When ingestion of antibiotics was followed by symptoms of vitamin B deficiency accompanied by degeneration of the coli flora, such vitamin deficiencies could be eliminated only by implantation of coli bacilli having properly functioning metabolic systems. The same vitamin deficiency, however, could not be removed solely by supplying the vitamin. It has been found that the implantation of intestinal microorganisms and particularly of lactobacillus acidophilus and L. bifidus is frequently beneficial. Some investigators have succeeded in substantially reducing the large number of dyspepsias observed after institutional antibiotic therapy by treatment aiming at substituting the destroyed coli bacilli.

A rise in hemoglobin level after oral ingestion of E. coli in iron-deficiency anemias has been reported and it has been pointed out that an abnormal bacterial flora which may be the responsible focus for disseminated toxic symptoms may frequently be overlooked in typical diseases of old age such as arthroses. In such cases it is recommended to implant a new non-pathogenic flora of physiologically active bacteria. The use of acidophilus and of E. coli after extended antibiotic treatment has been recommended but it has also been emphasized that E. coli established in the stomach or in the duodenum constitutes an additional focus of disease. Some investigators consider oral application of liquid coli cultures dangerous inasmuch as large portions of the ingested germs are destroyed by the hydrochloric acid of the gastric juices (pH 1.2—1.7) and therefore ingestion of living bacteria in capsules soluble in the intestinal fluid has been recommended.

The physiologically normal intestinal flora of humans is composed primarily of three symbionts: lactobacillus acidophilus, lacto-

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5 bacillus bifidus and escherichia coli. Until now,  
these three microorganisms were therapeutic-  
ally employed in separate doses since simul-  
taneous use of the three germs was not possi-  
10 ble for technical reasons unless a very com-  
plicated method was employed, namely of  
having three types of bacteria in a single pre-  
paration, which was not practically possible  
under the conditions of manufacture of a  
15 therapeutic preparation under industrial con-  
ditions.

The method of ingestion is of decisive im-  
portance in any bacterial substitution therapy.  
It has been found that bacteria given by  
15 mouth, for example, in liquid-suspended  
form, in tablets, in granules, or as powders,  
are already attacked by hydrochloric acid in  
the gastric juices and that the bacteria may  
from the stomach reach parts of the body  
20 where they may have pathogenic effects, such  
as E.coli in the gall bladder. It has been re-  
ported that of seventy patients who received  
E.coli by mouth in the form of the liquid  
culture, ten developed infections of the bile  
25 ducts. Such side effects may be avoided by  
employing capsules which resist gastric juices  
and dissolve in the intestinal fluid. It has  
been found, however, that bacteria filled into  
capsules by conventional methods lose their  
30 vital functions after a short storage time. This  
is due primarily to the fact that moisture is  
generally harmful. This not only holds for the  
usual bacteria preparations in paste form.  
Even the dry bacterial preparations produced  
35 according to several known methods in the  
form of powders or as granules soon lose their  
efficiency when filled into capsules since even  
the small amount of moisture contained in the  
capsule material and the atmospheric mois-  
40 ture penetrating through the capsules are  
detrimental to the survival rate of the bac-  
teria. The bacterial titer of such capsules  
which were not treated by the method of the  
present invention dropped from one billion to  
45 0 within a matter of days when stored at room  
temperature, and within a few weeks when  
stored under refrigeration at 4° C. Accord-  
ingly such preparations are not commercially  
suitable since it is not possible to keep pre-  
50 parations alive during the commercially re-  
quired periods of up to one year or longer,  
whether the preparations be stored under re-  
frigeration or at room temperature.

It is known to store microorganisms which  
55 were grown in liquid or on solid nutrient  
media under suitable conditions of growth,  
which were then separated from the media by  
centrifugation, filtration, or decantation, and  
which were finally lyophilized, in the form of  
60 a dry powder under vacuum or in a sealed  
glass vial filled with nitrogen. Depending on  
the method of preparation, the ratio of surviv-  
ing organisms still drops more or less rapidly.  
Such lyophilisates are not only used thera-  
65apeutically as such, but they also serve for the

production of fresh cultures. For this purpose  
the contents of the vial are added to a nutrient  
bouillon. A fresh bacterial culture is thus  
obtained and has to be taken by the patient.  
70 Bacterial cultures in such an ingestible form  
are objectionable from an esthetic point of  
view and provoke the patient's disgust since  
the liquid acquires an evil smell after a very  
short time. Furthermore, they have the dis-  
75 advantage of being free in the stomach when  
they can be attacked by acids and from which  
they can travel to undesired locations.

In order to employ several types of bacteria  
simultaneously as required in intestinal  
therapy, it would be necessary to inoculate a  
80 plurality of as many different culture fluids  
as there are types of bacteria which are to be  
applied in the intended therapeutic process.  
This would make it necessary to obtain each  
species of bacteria individually in dry form  
85 and to store it in an individual vial which is  
then used in the above-described manner for  
the preparation of an individual fresh culture.  
It is not possible to cultivate a plurality of  
different dry bacterial preparations simul-  
90 taneously in the same nutrient medium be-  
cause the several species would mutually inter-  
fere in their development or may require  
specific different nutrient substrates. Aside  
from the inconvenience of first preparing fresh  
95 cultures from dry bacterial preparations, this  
form of ingestion is objectionable primarily  
because fresh liquid bacterial cultures have to  
pass through the stomach and the duodenum  
and may be partially killed there by the  
100 hydrochloric acid in the gastric juices, or they  
may have harmful side effects.

It is accordingly a primary object of the  
present invention to provide for the produc-  
tion of living microorganism preparations, or  
105 at least preparations which have the capa-  
bility of again living, and which can remain  
stable over prolonged periods of time, can be  
taken after such long period of time and in  
the body can become alive and have the de-  
110 sired effect.

It is another object of the present invention  
to provide living microorganism preparations  
in capsules which are soluble in the intestine  
115 so that upon ingestion of the capsules the  
same do not become dissolved until they  
reach the intestine and the bacteria are not  
freed for action until they reach the intestine.  
This avoids the disadvantage of microorgan-  
isms becoming freed or free in the stomach  
120 where they can be attacked by acid, and also  
from where they can pass to undesired loca-  
tions such as the gall bladder.

It is still another object of the present in-  
vention to provide a method of preparing in-  
125 gestible dry microorganism preparations in  
intestinally soluble capsules.

Other objects and advantages of the pre-  
sent invention will be apparent from a further  
reading of the specification and of the 130

microorg. - lyophilized -> dry powder  
vacuum / N<sub>2</sub>

appended claims.

With the above objects in view, the present invention mainly comprises a storable live microorganism preparation which can be directly ingested and freed in the intestine of the person ingesting the same, comprising a dry enteric capsule containing dry, lyophilized living microorganisms of the intestinal flora, and a sealed moisture-proof container enclosing the capsule.

The arrangement for storing living microorganisms in accordance with the present invention mainly comprises a dry enteric capsule containing dry, lyophilized living microorganisms, and a sealed moisture-proof container enclosing the capsule.

In accordance with the present invention it is possible to avoid the disadvantage of bacterial preparations in enteric capsules, that is capsules that are specially treated so as to pass through the stomach and be dissolved in the intestinal juices, whereby the killing off of the bacteria in the capsule is to a great extent avoided so that the capsules can be stored for prolonged periods of time, for example 18 months or more, and used when desired without any substantial loss of the original action.

In accordance with the present invention it is preferred to utilize as the microorganism the microorganisms of the intestinal flora, for example *lactobacillus acidophilus*, *lactobacillus bifidus* or *escherichia coli*. Furthermore in accordance with the present invention it is possible to utilize not only one type of microorganism, but a plurality, and even all three of these different types of microorganisms.

It is further preferred in accordance with the present invention to include together with the dry lyophilized living microorganisms a non-toxic water absorbing substance, such as lactose anhydride, silica gel, Aerosol, activated carbon, aluminum, oxide, aluminum hydroxide, cellulose, tylose, magnesium carbonate, magnesium hydroxide, magnesium oxide, magnesium trisilicate, alkaline earth metal carbonates, oxides and hydroxides, kieselguhr, bentonite or barite.

It is still further preferred in accordance with the present invention to include along with the dry lyophilized living microorganisms a dry substance which is adapted in the presence of moisture, i.e. in the intestinal fluid after the capsule becomes dissolved, to act as a nutrient for the microorganisms, for example, lactose, dextrose, powdered skimmed milk, ascorbic acid, tylose, liver extract, yeast extract, powdered placenta, or the like. The capsules which are utilized for the purposes of the present invention may be any enteric capsules, that is capsules which pass through the stomach and become dissolved in the intestine. For example, specially treated gelatin capsules are particularly suitable. The capsule itself is predried prior to being filled with

the microorganisms, preferably to a water content of 1% by weight or less. The microorganisms which are dried are preferably dried to a water content of between 0.1 to 1% by weight, and most preferably to a water content of about 0.5% by weight.

In order to protect the microorganisms in the gelatin capsules from coming in contact with excessive moisture during prolonged storage, which would cause the microorganisms to grow in number in the capsule and subsequently to kill themselves off, the capsule containing the microorganism should be placed in a moisture-proof container, for example an evacuated, sealed metal foil container. the capsules may be enclosed individually or in number in a container.

The bacterial powder, that is the dry lyophilized microorganism preparation may be prepared as follows:—

A sterilized nutrient solution, for example 1% of glucose bouillon, is inoculated with *lactobacillus acidophilus* and is incubated at 37° C for 48 hours. The liquid is then centrifuged and the moist mass of bacteria is provided in known manner with preservatives and is lyophilized. It has been found that the kind and amount of preservatives added can help to keep the bacillus alive over prolonged periods of time. Thus, for example, utilizing 2 g of lactose, 2 g of skimmed milk powder and 40 g of ascorbic acid to 3 g of moist centrifuged bacterial substance, an initial titer of two billion will result in a survival of thirteen million germs after one year's storage at room temperature.

The bacterial substance is made into a powder by freeze-drying and is moved under exclusion of air and moisture (by working in a dry atmosphere of nitrogen or carbon dioxide) with moisture absorbing substances such as lactose anhydride, silica gel, or the like, and is filled into capsules which are soluble in the intestinal fluid and which capsules have first been predried, for example over phosphorous pentoxide. After additional drying, the filled capsules are transferred to preferably evacuated storage containers.

Experiments have shown that lyophilized bacteria which were filled into insufficiently predried gelatin capsules, that is capsules which still contain water, remain alive only for a very short period of time and that it is a necessary prerequisite for a sufficiently high rate of survival that moisture be removed to a very great extent not only from the bacteria, but also from the capsules.

In a comparative experiment, small undried gelatin rings were slipped over some of a number of predried gelatin capsules containing lyophilized bacteria. After six months to a year of storage it was found that several million of living bacteria could be detected only in those capsules which were completely dried and which had not been equipped with

Filled into capsules

undried gelatin rings.

It is a particular advantage of the present invention that it makes it possible to mix several types of bacteria in powder form and to maintain them simultaneously alive in a single capsule soluble in intestinal fluid, i.e. an enteric capsule, since they do not interfere with each other in the dry condition in which they are stored. It is possible, for example, to preserve several types of bacteria, for example lactobacillus acidophilus, lactobacillus bifidus, and escherichia coli in a single capsule and to show the presence of all three types of bacteria on a nutrient blade after storage of the capsule for one year or more. This shows that different bacteria will remain alive together in the above described powder form without damage, in an amount necessary for therapeutic effect over an adequate span of time.

It has been found that suitable preservatives for the present invention to prevent the individual cells of the microorganisms from being damaged or crushed during freezing which causes formation of ice crystals and which would otherwise crush the microorganisms include, as mentioned above, lactose, tylose, ascorbic acid, placenta powder, liver extract or yeast extract. When the bacteria which were kept alive but whose metabolism has been stopped so that they were, after a fashion, mummified, reached the intestinal tract, these preservatives created in the intestine favorable physiological conditions for the bacteria which caused the bacteria to resume metabolism and to multiply in the moisture and warmth prevailing in the intestine. It is also preferred to add to the otherwise finished dry preparation dry nutrient additives so that the bacteria when again provided with moisture in the intestinal canal immediately find substances which enhance their development. As mentioned above, dry milk powder or the

like may be used for this purpose.

Filling of the capsules may further be facilitated and ready dispersion of the preparations in the intestine may be achieved by making a paste from the powdery lyophilisates with anhydrous liquids such as paraffin oil, and by filling the resulting paste into capsules. Experiments have shown that lyophilized bacteria mixed with paraffin oil had a survival rate after one year of storage of 65 millions of coli bacilli and 10 millions of lactobacillus acidophilus per capsule.

It is particularly advantageous to store the filled capsules in such a manner that several filled capsules are sealed between two foils impervious to air and moisture which are welded together along their projecting edges, a vacuum preferably being formed in the storage space between the foils. Storing capsules containing living bacteria in glass vials is always disadvantageous since the vials may splinter during opening so that glass splinters may be ingested together with the capsule.

A normal unit dose of the bacteria per patient would be about 2 billion of the various types of bacteria. Of course this amount may vary greatly depending upon the individual patient, the individual need, and the type of bacteria. The capsules of the present invention may contain practically any amount of bacteria, for example from one half million to two billion or even more. The greater the number of bacteria per capsule the less the number of capsules that have to be taken.

The amount of protective substances which is mixed with the microorganisms prior to freeze-drying thereof may vary depending upon the substance. The following table will indicate preferred amounts, the amounts indicated in the table being with respect to 30 g of centrifuged wet bacteria mass:

TABLE.

	Maximum	Minimum	Optimum
Lactose	30 g	1 g	5 g
Dextrose	30 g	1	5
Powdered skimmed milk	30	5	10
Ascorbic Acid	0.75	0.075	0.4
Tylose	0.75	0.075	0.4
Liver Extract	5	0.25	1.5
Yeast Extract	5	0.25	1.5
Placenta powder	10	1	7

The amount of the moisture absorbing substances mixed with the lyophilisates may also vary depending upon the type of substance and the bacteria. Thus, for example, silica gel may be used in an amount of 1 to 60 g, most preferably 15 g per each 30 g of moist bacteria mass, while substances such as lactose anhydride, Aerosol, activated carbon, aluminum oxide, aluminum hydroxide, cellu-

lose, tylose or other cellulose derivatives, magnesium carbonate, magnesium oxide, magnesium hydroxide, magnesium trisilicate, alkaline earth metal carbonates, oxides and hydroxides, kieselguhr, barite and bentonite may be used in the same amounts. Lactose anhydride is preferably used in an amount of 5 to 30 g, most preferably 10 g per each 30 g of moist bacteria mass.

The following Examples are given to further illustrate the present invention. The scope of the invention is not, however, meant to be limited to the specific details of the Examples:

#### EXAMPLE 1.

24 litres of mash bouillon are inoculated with *Lactobacillus acidophilus*, are incubated 24 hours at 37° C and are then centrifuged. 30 g of a moist bacterial substance are obtained which contains about 5 g of dry bacteria and 25 g of water, 10 g skim milk powder, 5 g lactose and 0.5 g tylose are added to the moist substance and are mixed thoroughly. The mixture is lyophilized in known manner (by freeze-drying in a vacuum), and about 20 g of a lyophilisate are obtained which has a net moisture content of less than 0.5%. This lyophilisate is mixed under exclusion of air and moisture (by working in an atmosphere of dry nitrogen or carbon dioxide) with 10 g silica gel which absorbs the afore-mentioned residual moisture. The mixture is filled into gelatin capsules under exclusion of air and moisture. The capsules are of standard size No. 4 and have an internal diameter of 4.6 mm, an external diameter of 5.0 mm and a length of 14.5 mm. Such gelatin capsules, when in equilibrium with normal room atmosphere have a moisture content of 12–16% and are therefore not suitable as enclosure of the dry mixture without further preparation. The gelatin capsules are therefore predried in a vacuum desiccator over phosphorous pentoxide until they have only a residual moisture content of about 1%. Taking into account certain unavoidable material losses during filling of the mixture into capsules, about 500 capsules of 0.06 g each may be obtained from the above mixture. The filled capsules are subjected to supplemental drying (in a vacuum desiccator over  $P_2O_5$ ).

The capsules are stored either in tightly closed glass containers (with ground glass stoppers) which are evacuated or filled with dry nitrogen, or several capsules are sealed between two foils impervious to air and moisture, the projecting edges of the foils being welded together in known manner.

#### EXAMPLE 2.

10 litres meat bouillon are inoculated with *Escherichia coli*, are incubated 24 hours at 37° C and are then centrifuged. About 30 g of a moist bacterial substance are obtained and are mixed with 20 g dextrose, 20 g skim milk powder, 1.5 g liver extract and 0.4 g ascorbic acid. The mixture obtained is then lyophilized. The resulting lyophilisate is mixed with 40 g lactose anhydride and the mixture is filled and stored as described in Example 1.

#### EXAMPLE 3.

Equal or approximately equal amounts of the dry preparations obtained by the pro-

cesses of Examples 1 and 2 are mixed and simultaneously a third dry preparation is added which was obtained in an analogous manner from *Lactobacillus bifidus*. The total mixture is filled into capsules and stored as indicated in Example 1.

#### WHAT I CLAIM IS:—

1. An arrangement for storing living microorganisms in a form in which the same can be directly ingested and freed in the intestine of the person ingesting the same comprising, in combination, a dry enteric capsule containing dry, lyophilized living microorganisms; and a sealed moisture-proof container enclosing said capsule.

2. An arrangement as claimed in Claim 1, wherein the dry enteric capsule has a maximum water content of 1% by weight.

3. An arrangement as claimed in Claim 1 or 2, wherein the dry enteric capsule is made of gelatin.

4. A storable live microorganism preparation which can be directly ingested and freed in the intestine of the person ingesting the same, comprising a dry enteric capsule containing dry, lyophilized living microorganisms of the intestinal flora, and a sealed moisture-proof container enclosing said capsule.

5. A storable live microorganism preparation as claimed in Claim 4, wherein the dry, lyophilized living microorganism of the intestinal flora is *Lactobacillus acidophilus*, *Lactobacillus bifidus* or *Escherichia coli*.

6. A storable live microorganism preparation as claimed in Claim 4 or 5, wherein at least one non-toxic water absorbing substance is added to the microorganisms.

7. A storable live microorganism preparation as claimed in Claim 6, wherein the non-toxic water absorbing substance is lactose anhydride, silica gel, Aerosol, activated carbon, aluminum oxide, aluminum hydroxide, cellulose, tylose, magnesium carbonate, magnesium hydroxide, magnesium oxide, magnesium trisilicate, alkaline earth metal carbonates, oxides and hydroxides, keiselguhr, bentonite or barite.

8. A storable live microorganism preparation as claimed in any of claims 4 to 7, wherein the capsule contains a plurality of different dry lyophilized living microorganisms of the intestinal flora.

9. A storable live microorganism preparation as claimed in any of Claims 4 to 8, wherein at least one dry substance adapted in the presence of moisture to act as a nutrient for said microorganisms is added.

10. A storable live microorganism preparation as claimed in Claim 9, wherein said dry substance adapted in the presence of moisture to act as a nutrient for said microorganisms is lactose, dextrose, powdered skimmed milk, ascorbic acid, tylose, liver extract, yeast extract or powdered placenta.

11. A storable live microorganism prepara-

tion as claimed in any of Claims 4 to 10, wherein the dry, lyophilized living microorganisms of the intestinal flora have a moisture content of from 0.1 to 1% by weight.

5 12. A method of preparing a storable live microorganism preparation which can be directly ingested and in which the microorganisms are freed in the intestine of the person ingesting the same, comprising the  
10 steps of introducing dry, lyophilized living microorganisms into a dry enteric capsule; further drying said capsule containing said microorganisms; and enclosing said dry enteric capsule containing said microorganisms  
15 in a moisture-proof container.

13. A method as claimed in Claim 12, wherein the dry, lyophilized living microorganisms having a moisture content of 0.1 to 1% by weight are introduced into the dry  
20 enteric capsule while excluding atmospheric moisture.

14. A method as claimed in Claims 12 and 13, wherein the dry, lyophilized living microorganisms having a moisture content of 0.1  
25 to 1% by weight are introduced into a dry enteric capsule having a maximum moisture content of 1% by weight under vacuum in the presence of phosphorous pentoxide

15. A method as claimed in any of Claims 12 to 14, wherein the dry enteric capsule containing said microorganisms is enclosed under vacuum in an evacuated metal foil  
30 moisture-proof container.

16. A method as claimed in any of Claims 12 to 15, wherein at least one non-toxic water absorbing substance is introduced into the capsule.  
35

17. A method as claimed in any one of Claims 12 to 16, wherein at least one dry substance adapted in the presence of water to act as a nutrient for said microorganisms is introduced into the capsule.  
40

18. A storable live microorganism preparation substantially as hereindescribed and with reference to any one of the foregoing Examples.  
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19. A method of preparing a storable live microorganism preparation substantially as hereindescribed and with reference to any one of the foregoing Examples.  
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H. A. L. VENNER,  
Chartered Patent Agent,  
1 Great James Street, Bedford Row,  
London, W.C.1.  
Agent for Applicant.